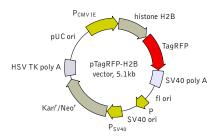


pTagRFP-H2B vector

The vector sequence has been compiled using the information from sequence databases, published literature, and other sources, together with partial sequences obtained by Evrogen. This vector has not been completely sequenced.



For vector sequence, please visit our Web site at http://www.evrogen.com/products/vectors.shtml

Product	Cat.#	Size	
pTagRFP-H2B vector	FP368	20 μ g	
Vector type	mammalian expression vector		
Reporter	TagRFP		
Reporter codon usage	mammalian		
Promoter for TagRFP	P _{CMV IE}		
Host cells	mammalian		
Selection	prokaryotic - kanamycin		
	eukaryotic - neomycin (G418)		
Replication	prokaryotic - pUC ori		
	eukaryotic - SV40 ori		
Use	red (orange) fluorescen	red (orange) fluorescent labeling of histone H2B	

Location of features

P_{CMV IE}: 1-589 Enhancer region: 59-465 TATA box: 554-560 Transcription start point: 583 Histone H2B: 657-1034 TagRFP: 1053-1766

SV40 early mRNA polyadenylation signal Polyadenylation signals: 1919-1924 & 1948-1953 mRNA 3' ends: 1957 & 1969

f1 single-strand DNA origin: 2016-2471 Bacterial promoter for expression of Kan^r gene -35 region: 2533-2538; -10 region: 2556-2561

Transcription start point: 2568 SV40 origin of replication: 2812-2947

SV40 early promoter Enhancer (72-bp tandem repeats): 2645-2716 & 2717-2788

2788 21-bp repeats: 2792-2812, 2813-2833 & 2835-2855 Early promoter element: 2868-2874

Major transcription start points: 2864, 2902, 2908 & 2913

Kanamycin/neomycin resistance gene Neomycin phosphotransferase coding sequences: Start codon (ATG): 2996-2998; Stop codon: 3788-3790 G->A mutation to remove Pst I site: 3178

C->A (Arg to Ser) mutation to remove BssH II site: 3524 Herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signal

Polyadenylation signals: 4026-4031 & 4039-4044 pUC plasmid replication origin: 4375-5018

Vector description

pTagRFP-H2B is a mammalian expression vector encoding TagRFP-H2B fusion protein. The vector can be used for fluorescent labeling of histone H2B in living cells.

TagRFP codon usage is optimized for high expression in mammalian cells (humanized) [Haas et al. 1996]. Human histone H2B is fused to the TagRFP N-terminus.

pTagRFP-H2B vector can be used as a source of TagRFP-H2B hybrid sequence. The vector backbone contains unique restriction sites that permit its excision and further insertion into expression vector of choice.

Note: The plasmid DNA was isolated from dam⁺-methylated *E.coli*. Therefore some restriction sites are blocked by methylation. If you wish to digest the vector using such sites you will need to transform the vector into a dam⁻ host and make fresh DNA.

The vector backbone contains immediate early promoter of cytomegalovirus ($P_{\text{CMV IE}}$) for protein expression, SV40 origin for replication in mammalian cells expressing SV40 T-antigen, pUC origin of replication for propagation in *E. coli*, and f1 origin for single-stranded DNA production. SV40 polyadenylation signals (SV40 poly A) direct proper processing of the 3'-end of the reporter mRNA.

SV40 early promoter (P_{SV40}) provides neomycin resistance gene (Neo^r) expression to select stably transfected eukaryotic cells using G418. Bacterial promoter (P) provides kanamycin resistance gene expression (Kan^r) in *E. coli*. Kan^r/Neo^r gene is linked with herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signals.

Expression in mammalian cells

pTagRFP-H2B vector can be transfected into mammalian cells by any known transfection method. CMV promoter provides strong, constitutive expression of the TagRFP-H2B fusion in eukaryotic cells. If required, stable transformants can be selected using G418 [Gorman 1985].

Propagation in E. coli

Suitable host strains for propagation in *E. coli* include DH5alpha, HB101, XL1-Blue, and other general purpose strains. Plasmid incompatibility group is pMB1/ColE1. The vector confers resistance to kanamycin (30 μ g/ml) to *E. coli* hosts. Copy number in *E. coli* is about 500.

References

Gorman, C. (1985). "High efficiency gene transfer into mammalian cells." In: DNA cloning: A Practical Approach, Vol. II. Ed. by Glover. (IRL Press, Oxford, U.K.) Pp. 143–190.

Haas, J. et al. (1996) "Codon usage limitation in the expression of HIV-1 envelope glycoprotein." Curr Biol, 6 (3): 315-324 / pmid: 8805248

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