

Anti-PhiYFP antibody

Cat. #AB601

Description

Rabbit polyclonal antibody against denatured TurboYFP and Phi-Yellow proteins.

Size: 100 µg.

Immunogen: Full-length recombinant non-denatured PhiYFP comprising 6XHis tag.

Preparation: Full-length recombinant PhiYFP comprising 6XHis tag was purified from transformed E. coli using metal-ion affinity chromatography. Antibody was produced in rabbits immunized with the recombinant non-denatured PhiYFP. Specific IgG were purified by PhiYFP affinity chromatography.

Formulation: Lyophilized from the buffer containing 0.1% mannitol, 0.1% dextran, 0.1M NaCl, 0.01M Na₂PO₄, 0.01M NaBO₄, pH 7.4.

Reconstitution: Reconstitute with 100 µl of sterile water or 50% glycerol.

Storage: Lyophilized samples are stable for twelve months from date of receipt when stored at -20°C or -70°C. The presence of silica gel drier is advisable.

Reconstituted with sterile water, antibody can be stored at +4°C for three months without detectable loss of activity.

Reconstituted with 50% glycerol, antibody can be stored at -20°C in a manual defrost freezer for six months without detectable loss of activity. Aliquot antibody upon reconstitution.

Avoid repeated freeze / thaw cycles.

Specificity: The antibody has been selected to recognize non-denatured TurboYFP, PhiYFP, and PhiYFP-m.

Note: *Heat- or chemically denatured proteins lack antigen determinants.*

The antibody shows little or no cross-reactivity with other fluorescent proteins like EGFP, TurboGFP, KFP-Red, and DsRed2.

Applications: immunoblotting, immunohistochemistry, ELISA, In cell Western, immunoprecipitation.

Recommendations for use

Working concentrations: For immunoblotting and immunohistochemistry use at a dilution of 1:20 000

For ELISA: use at a dilution of 1:20 000 - 1:30 000

Note: Optimal dilutions/concentrations should be determined by the end user.

Tissue (cells) fixation for immunohistochemistry: Formaldehyde (formalin, paraform) fixation is recommended because it does not cause antigenicity loss. Do not use any protein-denaturing agents like glutaraldehydes, alcohols, or picric acid. For example, tissues can be fixed in PBS containing 4% formaldehyde for 10-15 min, treated with 0.1% saponin in PBS for 10-15 min, and washed three times in PBS.

Sample preparation for immunoblotting: Use a non-denaturing buffer for tissue homogenization. Treat the sample by ultrasound to cut genomic DNA (2-3 impulses of minimal power is enough for a sample of 50 μ l).

To a sample containing 1-100 ng of a target protein, add an equal volume of 2x SDS-PAGE sample buffer.

Note: Do not heat the samples before loading on a gel or spotting on a membrane (for dots).

Note: PAAG mobility of non-denatured proteins differs that of from denatured ones and often does not reflect protein molecular weight. Usually, immunostaining results in one or more diffuse bands corresponding to a non-denatured and a partially denatured protein.

Notice to Purchaser:

This product is intended for research use only.

MATERIAL SAFETY DATA SHEET INFORMATION: To the best of our knowledge, these products do not require a Material Safety Data Sheet. However, all the properties of these products (and, if applicable, each of their components) have not been thoroughly investigated. Therefore, we recommend that you use gloves and eye protection, and wear a laboratory coat when working with these products.