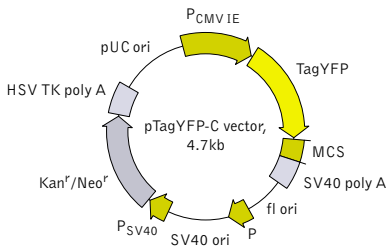


## pTagYFP-C vector

The vector sequence has been compiled using the information from sequence databases, published literature, and other sources, together with partial sequences obtained by Evrogen. This vector has not been completely sequenced.



For vector sequence, please visit our Web site at <http://www.evrogen.com/support/vector-info.shtml>

### pTagYFP-C vector MCS

$\xrightarrow{\text{TagYFP}}$ 
 $\xrightarrow{\text{BspE I}}$ 
 $\xrightarrow{\text{Xho I}}$ 
 $\xrightarrow{\text{Hind III}}$ 
 $\xrightarrow{\text{Pst I}^*}$ 
 $\xrightarrow{\text{Kpn I}}$ 
 $\xrightarrow{\text{Apa I}}$ 
 $\xrightarrow{\text{BamH I}}$ 
 $\xrightarrow{\text{STOPs}}$ 
  
 . . . TCC . GGA . CTC . AGA . TCT . CGA . GCT . CAA . GCT . TCG . AAT . TCT . GCA . GTC . GAC . GGT . ACC . GCG . GGC . CCG . GGA . TCC . ACC . GGA . TCT . AGA . TAA . CTG . ATC . A . . .

\* — not unique sites.

# — sites are blocked by *dam* methylation. If you wish to digest the vector with these enzymes, you will need to transform the vector into a *dam*<sup>-</sup> host and make fresh DNA.

### Location of features

**P<sub>CMV IE</sub>:** 1-589  
**Enhancer region:** 59-465  
**TATA box:** 554-560  
**Transcription start point:** 583  
**TagYFP**  
**Kozak consensus translation initiation site:** 606-616  
**Start codon (ATG):** 613-615; **Stop codon:** 1408-1410  
**Last amino acid in TagYFP:** 1327-1329  
**MCS:** 1330-1407  
**SV40 early mRNA polyadenylation signal**  
**Polyadenylation signals:** 1550-1555 & 1579-1584  
**mRNA 3' ends:** 1588 & 1600  
**f1 single-strand DNA origin:** 1647-2102  
**Eukaryotic promoter for expression of Kan<sup>r</sup> gene**  
**-35 region:** 2164-2169; **-10 region:** 2187-2192  
**Transcription start point:** 2199  
**SV40 origin of replication:** 2443-2578  
**SV40 early promoter**  
**Enhancer (72-bp tandem repeats):** 2276-2347 & 2348-2419  
**21-bp repeats:** 2423-2443, 2444-2464 & 2466-2486  
**Early promoter element:** 2499-2505  
**Major transcription start points:** 2495, 2533, 2539 & 2544  
**Kanamycin/neomycin resistance gene**  
**Neomycin phosphotransferase coding sequences:**  
**Start codon (ATG):** 2627-2629; **Stop codon:** 3419-3421  
**G->A mutation to remove Pst I site:** 2809  
**C->A (Arg to Ser) mutation to remove BssH II site:** 3155  
**Herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signal**  
**Polyadenylation signals:** 3657-3662 & 3670-3675  
**pUC plasmid replication origin:** 4006-4649

### References

Gorman (1985). "High efficiency gene transfer into mammalian cells." In: *DNA cloning: A Practical Approach, Vol. II*. Ed. by Glover. (IRL Press, Oxford, U.K.) Pp. 143–90.  
 Haas et al. (1996) "Codon usage limitation in the expression of HIV-1 envelope glycoprotein." *Curr Biol*, 6 (3): 315–24 / PMID: 8805248  
 Kozak (1987) "An analysis of 5'-noncoding sequences from 699 vertebrate messenger RNAs." *Nucleic Acids Res*, 15 (20): 8125–48 / PMID: 3313277

Product	Cat.#	Size
pTagYFP-C vector	FP131	20 µg
The price does not include delivery. The price varies in different countries. Please contact your local distributor for exact prices and delivery information.		
Vector type	mammalian expression vector	
Reporter	TagYFP	
Reporter codon usage	mammalian	
Promoter for TagYFP	P <sub>CMV IE</sub>	
Host cells	mammalian	
Selection	prokaryotic - kanamycin eukaryotic - neomycin (G418)	
Replication	prokaryotic - pUC ori eukaryotic - SV40 ori	
Use	TagYFP expression in mammalian cells; generation of fusions to the TagYFP C-terminus	

### Vector description

pTagYFP-C is a mammalian expression vector encoding yellow fluorescent protein TagYFP. The vector allows generation of fusions to the TagYFP C-terminus and expression of TagYFP fusions or TagYFP alone in eukaryotic (mammalian) cells.

TagYFP codon usage is optimized for high expression in mammalian cells (humanized) [Haas et al. 1996]. To increase mRNA translation efficiency, Kozak consensus translation initiation site is generated upstream of the TagYFP sequence [Kozak 1987]. Multiple cloning site (MCS) is located between TagYFP coding sequence and SV40 polyadenylation signal (SV40 polyA).

The vector backbone contains immediate early promoter of cytomegalovirus (P<sub>CMV IE</sub>) for protein expression, SV40 origin for replication in mammalian cells expressing SV40 T-antigen, pUC origin of replication for propagation in *E. coli* and f1 origin for single-stranded DNA production. SV40 polyadenylation signals (SV40 poly A) direct proper processing of the 3'-end of the reporter mRNA.

SV40 early promoter (P<sub>SV40</sub>) provides neomycin resistance gene (Neo<sup>r</sup>) expression to select stably transfected eukaryotic cells using G418. Bacterial promoter (P) provides kanamycin resistance gene expression (Kan<sup>r</sup>) in *E. coli*. Kan<sup>r</sup>/Neo<sup>r</sup> gene is linked with herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signals.

### Generation of TagYFP-fusion proteins

A localization signal (or a gene of interest) should be cloned into MCS of the vector. It will be expressed as a fusion to the TagYFP C-terminus when inserted in the same reading frame as TagYFP and no intervening stop codons are present. TagYFP-tagged fusions retain fluorescent properties of the native protein allowing fusion localization *in vivo*. Unmodified vector will express TagYFP, when transfected into eukaryotic (mammalian) cells.

**Note:** The plasmid DNA was isolated from *dam*<sup>+</sup>-methylated *E. coli*. Therefore some restriction sites are blocked by methylation. If you wish to digest the vector using such sites you will need to transform the vector into a *dam*<sup>-</sup> host and make fresh DNA.

### Expression in mammalian cells

pTagYFP-C vector can be transfected into mammalian cells by any known transfection method. If required, stable transformants can be selected using G418 [Gorman 1985].

### Propagation in *E. coli*

Suitable host strains for propagation in *E. coli* include DH5alpha, HB101, XL1-Blue, and other general purpose strains. Plasmid incompatibility group is pMB1/ColE1. The vector confers resistance to kanamycin (30 µg/ml) to *E. coli* hosts. Copy number in *E. coli* is about 500.

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